

Chalcone-ligated molybdenum carbonyl complexes:  
Synthesis and evaluation as quadruplex DNA binders

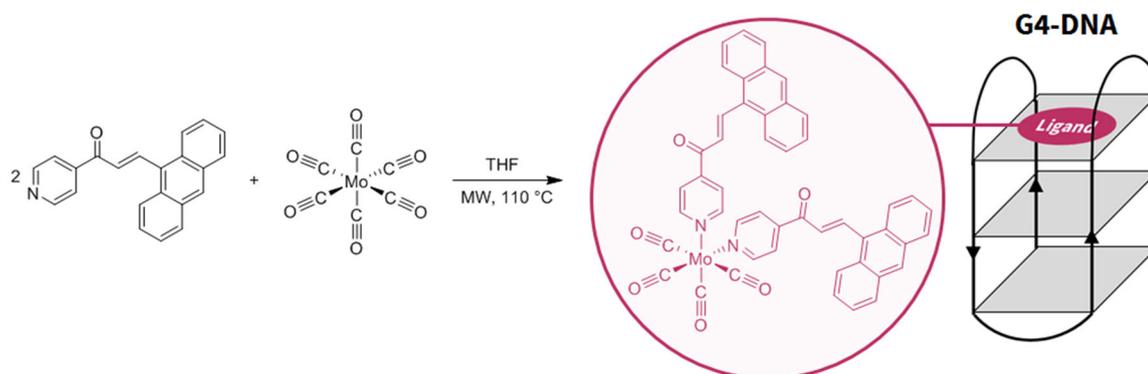
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**Abstract**

The microwave-assisted reaction of  $\text{Mo}(\text{CO})_6$  with isomeric pyridyl anthryl chalcones **1a-f** afforded six novel molybdenum carbonyl complexes **2a-f**. Complexes **2a-f** have been characterized by infrared and  $^1\text{H}$  Nuclear Magnetic Resonance spectroscopies, and the structures of **2a-d** were revealed to be *cis*-disubstituted complexes with general formula  $\text{Mo}(\text{CO})_4\text{L}_2$ . The intercalative abilities of **2a-d** towards quadruplex DNA were examined using the Fluorescent Intercalator Displacement (FID) assay.

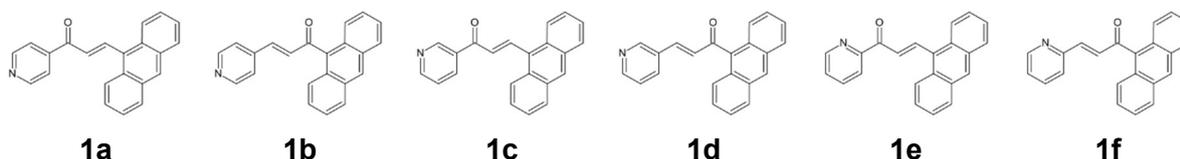


## 1. Introduction

Recently, a structure known as the G-quadruplex (G4) has emerged as a promising anti-cancer drug target. G4 structures are helical structures formed in guanine-rich regions of RNA or DNA sequences stabilised by sodium or potassium cations [1]. Such structures are found in higher frequency in telomeres and promoters [2], suggesting that they may play a role in cancer regulation. It is hence hypothesised that molecules that can stabilise G4 structures may display anti-cancer activity.

Chalcones are a class of plant secondary metabolites featuring the  $\alpha,\beta$ -unsaturated ketone pharmacophore and display a wide range of biological activity [3]. Pyridyl anthryl chalcones contain the planar anthryl group, which has a large  $\pi$  surface area, potentially allowing it to intercalate between the guanine bases and act as a ligand for G4 structures, enhancing its anti-cancer activity. On the other hand, molybdenum carbonyl complexes have been investigated as Carbon Monoxide Releasing Molecules (CORMs) which can release therapeutic amounts of carbon monoxide to tissues [4]. The attachment of pyridine anthryl chalcones to molybdenum carbonyl complexes may enhance its G4-binding ability due to electrostatic attraction between the metal center and the negatively-charged phosphate backbone of the G4-DNA.

In this work, molybdenum carbonyl complexes **2a-f** were synthesised by the reaction of  $\text{Mo}(\text{CO})_6$  with the anthryl pyridine chalcones **1a-f** (Chart 1). The intercalative abilities of **2a-d** towards quadruplex DNA were examined to ascertain their potential as anti-cancer agents.



**Chart 1.** Structures of **1a-f**

## 2. Experimental

### *Synthesis of molybdenum complexes 2a-f*

$\text{Mo}(\text{CO})_6$  (15 mg, 57  $\mu\text{mol}$ ) and the corresponding chalcone (2 molar equivalent) were added to 1 mL of tetrahydrofuran and reacted in a Discover-SP microwave reactor. The solvent was removed under reduced pressure and the residue was washed with hexane and air dried to yield the solid product.

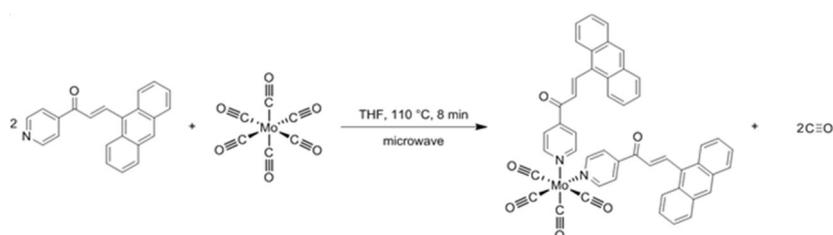
### *Fluorescent Intercalator Displacement (FID) assay*

The FID assay [5] was performed on **2a–d** against G4 quadruplex DNA (HTelo and *c-myc*) as well as duplex DNA (ds26) to obtain their corresponding half-maximal degradation concentration ( $DC_{50}$ ) values.

### 3. Results and Discussion

#### Synthesis and characterization

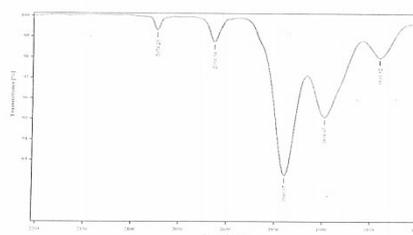
The reaction of  $Mo(CO)_6$  with the respective chalcone **1a–d** afforded the novel *cis*-disubstituted molybdenum carbonyl complexes **2a–d** (Scheme 1) in moderate yields. All complexes were characterised by infrared (IR) and  $^1H$  Nuclear Magnetic Resonance (NMR) spectroscopies.



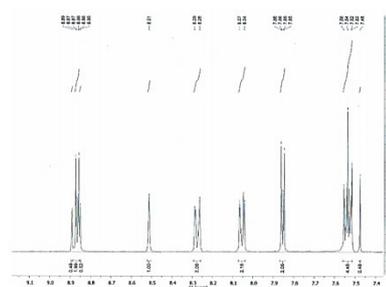
**Scheme 1.** Synthesis of the *cis*-disubstituted complex **2a**

The IR spectra of **2a–d** in the  $\nu_{CO}$  region (Figure 1) resembles those of previously reported *cis*- $Mo(CO)_4(amine)_2$  complexes [6], confirming the identities of **2a–d**.

The  $^1H$  NMR spectra of all complexes displayed one set of signals, suggesting that the complexes are symmetrical in solution. The assignment of protons on **2a** (Figure 2) is used as an example. The singlet at  $\delta = 8.51$  ppm can be assigned to the proton on position 10 on the anthracene ring, while the doublet at  $\delta = 8.87$  ppm can be assigned to the protons at the *ortho* position of the pyridine ring. The two sets of doublets at  $\delta = 8.87$  ppm and  $\delta = 7.54$  ppm with coupling constant  $J = 16.0$  Hz can be assigned to the vinylic protons. The value of the coupling constant suggests that the chalcone exists in the *trans* configuration.



**Figure 1.** IR spectrum of **2a** in dichloromethane solution



**Figure 2.**  $^1H$  NMR spectrum of **2a** in  $CDCl_3$

#### Fluorescent Intercalator Displacement (FID) Assay

The  $DC_{50}$  values of **2a–d** were determined using the FID assay (Table 1); a lower  $DC_{50}$  value is indicative of better quadruplex intercalative properties.

While  $Mo(CO)_6$  and **1a–d** did not bind to quadruplex DNA, complexes **2a–d** were able to intercalate to HTelo and *c-myc*. This suggests that there might be synergistic effects, such as

electrostatic attraction, which allow **2a-d** to intercalate. All four complexes showed greater selectivity for HTelo over *c-myc*. This might be because *c-myc* adopts a parallel topology while HTelo forms a (3+1) hybrid, and the more compact structure of *c-myc* would increase the difficulty of intercalation. However, all complexes showed

greatest selectivity for duplex DNA, suggesting an alternative mode of interaction besides intercalation.

#### 4. Conclusion

Six novel molybdenum carbonyl complexes with chalcone ligands were synthesized and characterized. FID assays revealed that **2a-d** displayed moderate intercalative abilities for quadruplex DNA while having greater selectivity for HTelo over *c-myc*.

#### 5. Future Work

The identities of **2e** and **2f** will be established followed by FID assays on these two complexes, and the brine shrimp lethality (BLT) assay will be carried out on complexes **2a-f** to evaluate their cytotoxicity.

#### 6. References

- [1] Howard, F. B.; Frazier, J.; Miles, H. T. *Biopolymers* **1977**, *16* (4), 791.
- [2] Chambers, V. S.; Marsico, G.; Boutell, J. M.; Di Antonio, M.; Smith, G. P.; Balasubramanian, S. *Nat. Biotechnol.* **2015**, *33* (8), 877.
- [3] K. Sahu, N.; S. Balbhadra, S.; Choudhary, J.; V. Kohli, D. *Curr. Med. Chem.* **2012**, *19* (2), 209.
- [4] Kromer, L.; Coelho, A. C.; Bento, I.; Marques, A. R.; Romão, C. C. *J. Organomet. Chem.* **2014**, *760*, 89.
- [5] Monchaud, D.; Allain, C.; Teulade-Fichou, M.-P. *Bioorg. Med. Chem. Lett.* **2006**, *16* (18), 4842.
- [6] Kraihanzel, C. S.; Cotton, F. A. *Inorg. Chem.* **1963**, *2* (3), 533.

Compound	DC <sub>50</sub> <sup>a</sup> / μM		
	HTelo	<i>c-myc</i>	ds26
<b>2a</b>	44.33 (2.29)	54.33 (1.82)	10.01 (0.18)
<b>2b</b>	39.87 (0.31)	74.77 (0.51)	9.03 (0.09)
<b>2c</b>	55.87 (1.22)	81.85 (3.03)	10.71 (0.34)
<b>2d</b>	39.20 (0.37)	102.88 (4.14)	10.35 (0.21)

<sup>a</sup> Standard errors (*N* = 3) are shown in parentheses.

**Table 1.** Affinity and selectivity of **2a-d** for oligonucleotides, determined by the FID assay